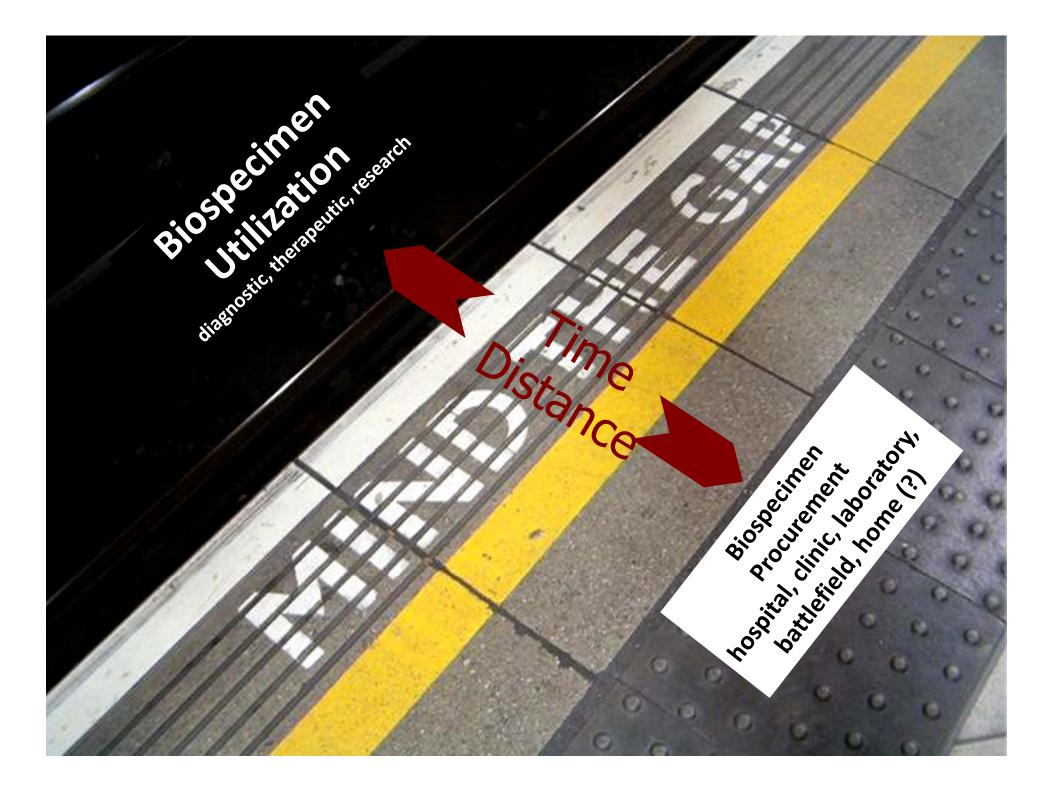


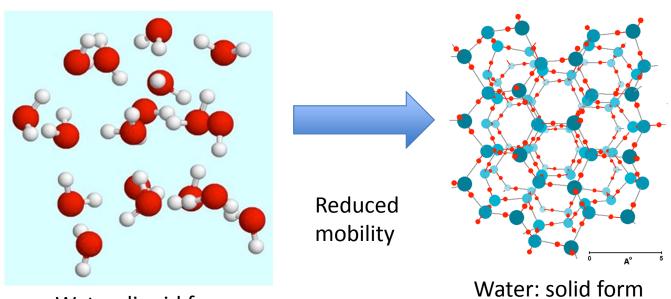
Using thermodynamic principles and practical realities to select a storage temperature

Allison Hubel, PhD
University of Minnesota





Changes in molecular mobility



Water: liquid form

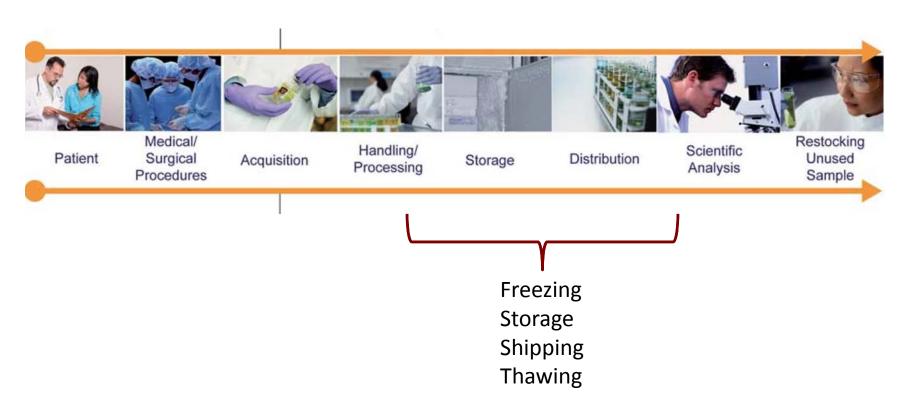
Protein

Proteases

Reduced rates of reaction
Reduced activity

University of Minnesota
Driven to Discover

Life-cycle of a biospecimen





Prefreeze processing Introduction of stabilizing agents **Cooling protocol** Storage conditions **Focus** Thawing/rewarming Post thaw assessment

Elements of a preservation protocol





Mechanical Freezers -20 C, -80 C, -153 C

Storage Options

Liquid nitrogen storage (-196 C)

- Liquid phase
- Vapor phase







The freezing process influences selection of the storage temperature



What do you know about storage of biospecimens because you have eaten a popsicle?

You can have solid and liquid together at the same time

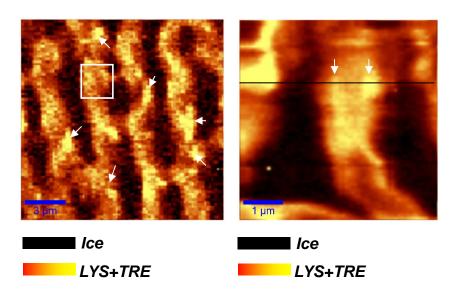
Solute is not incorporated into the ice

Complex mixtures freeze over a range of temperatures (not a single temperature)

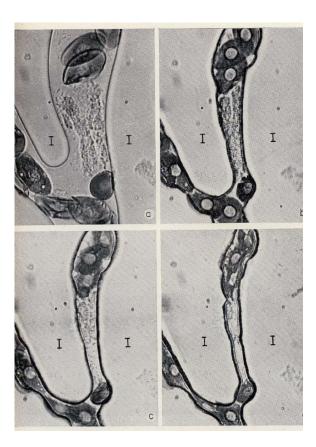




Partitioning of biospecimen during freezing

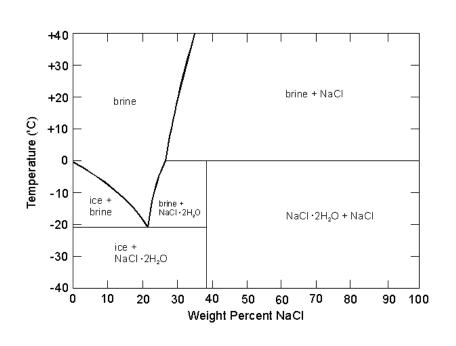


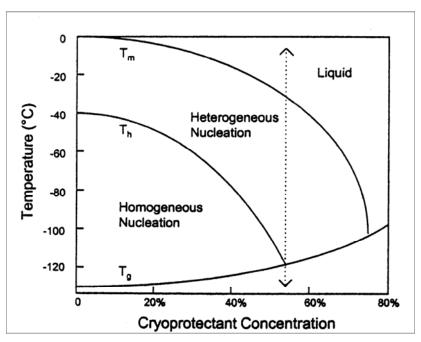
- Water is removed in the form of ice
- Proteins/cells partitioned into gap
- Size of gap decreases with decreasing temperature
- Concentration in gap increases with decreasing temperature





When does freezing end?





Option #1: eutectic

- Assumes that sample fully solidifies
- T_{eut}= -21 C (isotonic saline)

Option #2: glass transition temperature

- Assumes sample partially vitrifies
- $T_g = -120 C (10\% DMSO solution)$
- T_g= -132 C (pure water)



Summary: freezing of aqueous solutions

- Biospecimens are complex mixtures
- Complex mixtures freeze over a range of temperatures
- Specimen is partitioned into unfrozen solution
- Sample is not fully frozen until eutectic or glass transition temperature
- Store at a temperature where sample is fully frozen



What do I do if I do not know the phase diagram for my solution?

Option #1: Estimate glass transition temperature mathematically:

$$T_g(mixture) = T_{g1} (1-x) + T_{g2}x + k*x*(1-x)$$

T_g=glass transition temperature k=interaction parameter X=weight fraction of component



Options, cont.

Option #2: Check scientific literature for stability studies

 Literature review being published in Biopreservation and Biobanking in June

Option #3: Use your QA/QC program for continuous improvement

More on this later

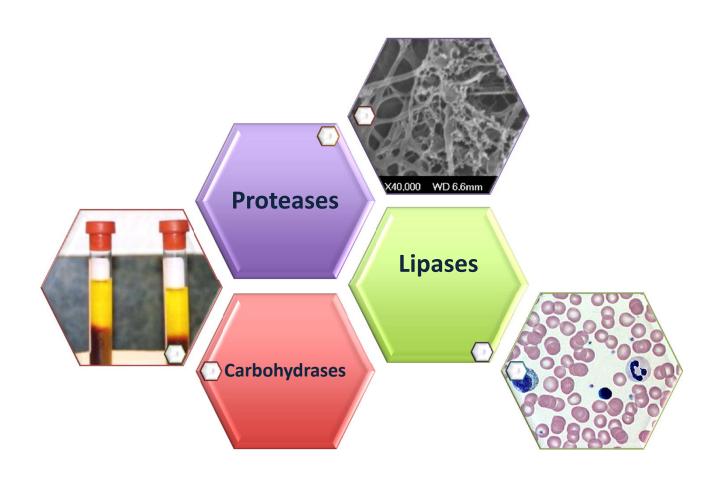




Selection of storage temperature: suppressing activity of degradative molecules



Degradative molecules are present in every sample





Activity of degradative molecules

Protein activity is a function of its dynamics



Reducing temperature reduces activity of molecule

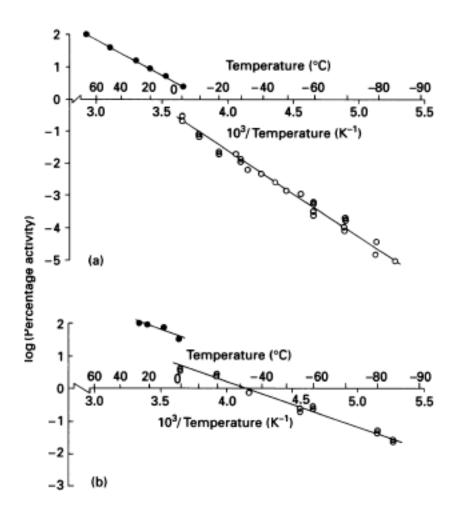
Activity can be described by Arrhenius Equation

$$k = A \cdot e^{-Ea/(R \cdot T)}$$

There is a threshold temperature below which no activity is observed



β-glucosiadase activity



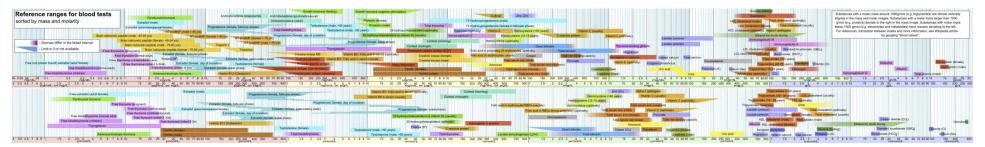
More, Biochem J, 1995

Enzymatic activity with temperature

- Reduced temperatures reduces activity of enzymes
- Measurable activity observed at low temperatures



Plasma Proteome



Low temperature activity

- Conventional wisdom: inactivation by ~-53 C for most proteins
 - > RNAase activity has been measured down to -93 C
 - \triangleright β -glucsoidase activity down to -70 C
- Low temperature activity of many enzymes have not been measured
- Samples should be stored at temperatures below which degradative molecules are inactivated





Factors that influence sample stability when in storage



Temperature Fluctuations

Result from accessing of working repositories

Amplitude and frequency vary with location in the repository and methods used for retrieving samples





Background ionizing radiation



This damage mechanism has been studied for cells. Not clear if other biospecimens experience the same



Summary

- Minimizing temperature fluctuations is also desirable
 - Training of biorepository workers
 - Controlling access to repository



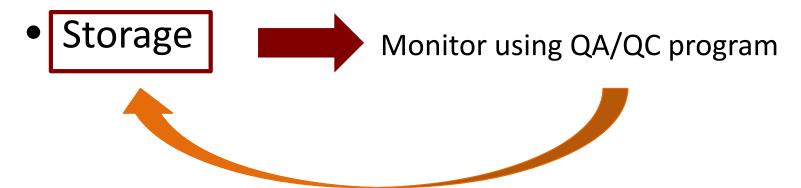


The role of QA/QC on selection of storage temperature and continuous improvement



Potential sources of degradation

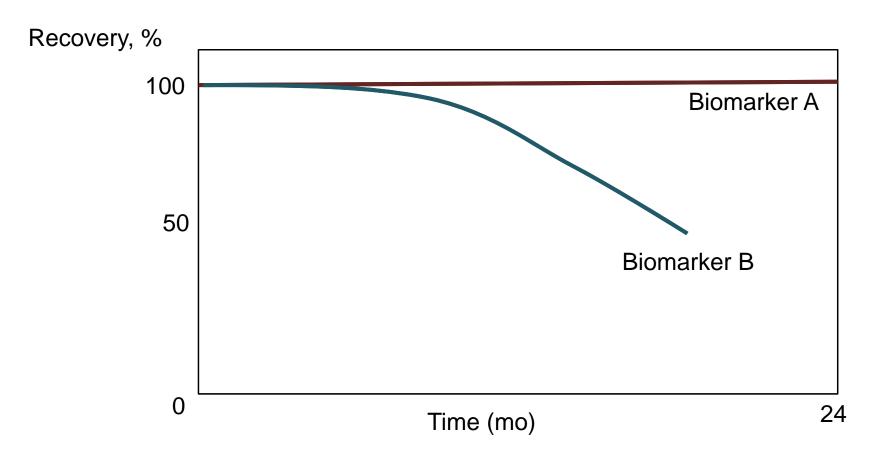
- Collection
- Extraction



Continuous improvement



Testing sample stability

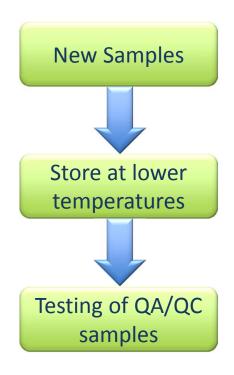


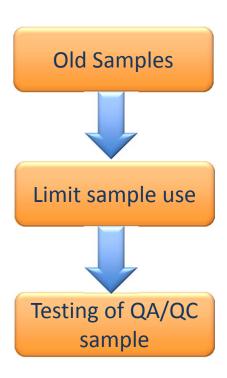
QA/QC program finds Biomarker B is not stable

> Samples are not stored correctly



Continuous Improvement







Upcoming article

Type: review-article

REVIEW ARTICLE

BIOPRESERVATION AND BIOBANKING Volume 12, Number 3, 2014 © Mary Ann Liebert, Inc. DOI:10.1089/bio.2013.0084

Storage of Human Biospecimens: Selection of the Optimal Storage Temperature

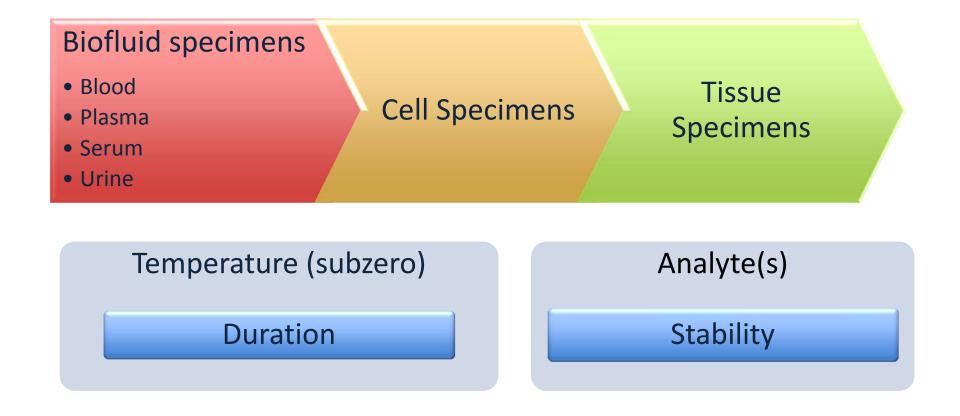
Allison Hubel, 1,2 Ralf Spindler, 1,2 and Amy P.N. Skubitz 1,3

Millions of biological samples are currently kept at low tempertures in cryobanks/biorepositories for long-term storage. The quality of the biospecimen when thawed, however, is not only determined by processing of the biospecimen but the storage conditions as well. The overall objective of this article is to describe the scientific basis for selecting a storage temperature for a biospecimen based on current scientific understanding. To that end, this article reviews some physical basics of the temperature, nucleation, and ice crystal growth present in biological samples stored at low temperatures (-20° C to -196° C), and our current understanding of the role of temperature on the activity of degradative molecules present in biospecimens. The scientific literature relevant to the stability of specific biomarkers in human fluid, cell, and tissue biospecimens is also summarized for the range of temperatures between -20° C to -196° C. These studies demonstrate the importance of storage temperature on the stability of critical biomarkers for fluid, cell, and tissue biospecimens.

➤ More complete discussion of these scientific principals



Literature review: storage stability studies



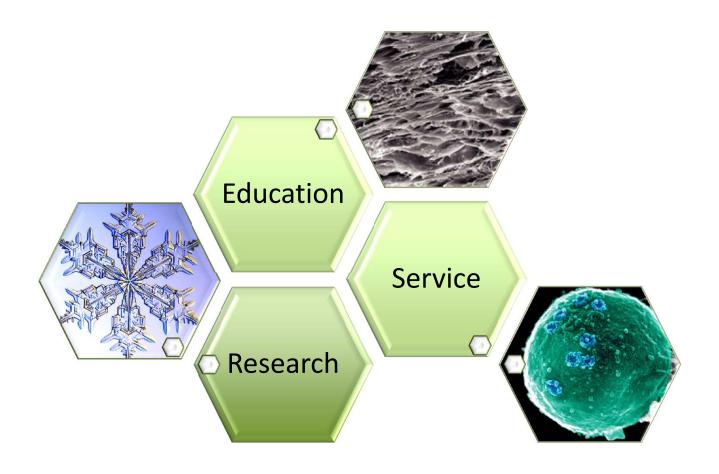


Summary

- Store at temperatures below which the sample is fully solidified
- Store at temperature below which the activity of all degradative molecules are inactive
- Verify stability



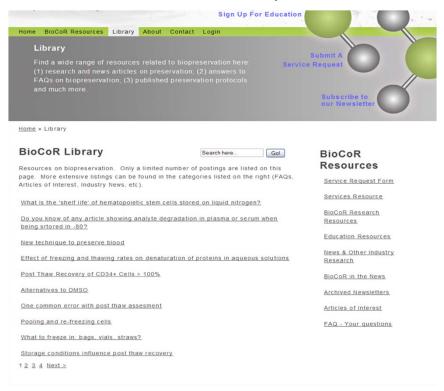
BioCoR Resources

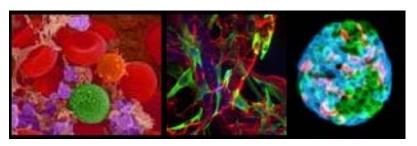




Education Resource

BioCoR library





Preservation of Molecular, Cellular and Tissue Biospecimens

Endorsed by ISBER



Preservation of Cellular Therapies

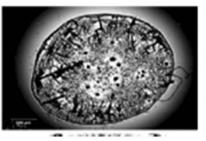


BioCoR Research

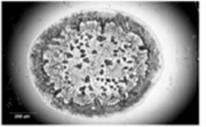
Dry State Storage of Plasma

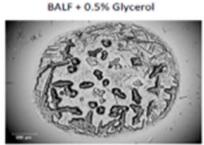
BALF + 0.1% Trehalose

Previously frozen, Dried at 0%RH



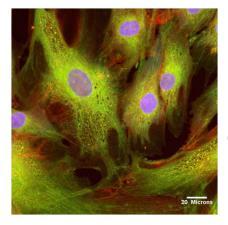
BALF only





sciencephotolibrary

QC tests for UCB



Improving preservation of MSCS

All projects are NIH funded





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<u>Co-authors</u> Amy Skubitz Ralf Spindler

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